CICH ₂ COOH	MW: 94.50	CAS: 79-11-8	RTECS: AF8575000
METHOD: 2008, Issue 2	EVA	LUATION: FULL	Issue 1: 15 May 1989 Issue 2: 15 August 1994
OSHA : no PEL NIOSH: no REL ACGIH: no TLV (1 ppm = 3.87 mg/m ³ @	NTP)	PROPERTIES:	solid; MP 61 to 63 °C; BP 189 °C; VP 0.1 kPa (0.75 mm Hg) @ 20 °C; flash point 126 °C; lower explosive limit in air 8% v/v

SYNONYMS: chloroethanoic acid; monochloroacetic acid

	SAMPLING	_	MEASUREMENT
SAMPLER:	SOLID SORBENT TUBE (silica gel, 100 mg/50 mg)	TECHNIQUE:	ION CHROMATOGRAPHY, CONDUCTIVITY DETECTION
FLOW RATE:	0.05 to 0.2 L/min	ANALYTE:	chloroacetate ion
VOL-MIN: -MAX:	1 L @ 0.25 ppm 100 L	DESORPTION:	2 mL deionized water
SHIPMENT:	routine	INJECTION LOOP VOLUME:	500 µL
SAMPLE		ELUENT:	1.5 mM NaHCO ₃ ; 1.0 mL/min
STABILITY: BLANKS:	at least 7 days @ 25 °C; 32 days refrigerated [1] 2 to 10 field blanks per set	COLUMNS:	Ion Pac AS4A Separator, AG4A Guard micromembranesuppressor(seeOTHER METHODS)
		CALIBRATION:	standard solutions of chloroacetic acid in deionized water
ACCURACY			
RANGE STUDIED:	0.35 to 29 mg/m ³ [1,2] (3-L samples)	RANGE:	1 to 80 μg per sample [1] : 0.04 μg per sample [2]
BIAS:	- 2.0%	PRECISION (Ŝ _r):	
OVERALL PRECISION (Ŝ _{rt}): 0.08 [1]			
ACCURACY:	± 17.7%		

APPLICABILITY: The working range is 0.09 to >85 ppm (0.3 to 30 mg/m³) for a 3-L air sample.

INTERFERENCES: Chloroacetyl chloride is a positive interferent since it is hydrolyzed to monochloroacetic acid by the measurement procedure and is efficiently collected by silica gel [3]. Particulate salts of the acid are positive interferents. The chromatographic conditions given will separate acetate, chloride, dichloroacetate, fluoride, glycolate, and trichloroace tate ions from chloroacetate ion.

OTHER METHODS: This revises P&CAM 332 [2]. The columns used in P&CAM 332 are no longer available. The newer columns indicated here show improvements in the analytical range and sensitivity.

2008

REAGENTS:

- 1. Water, filtered, deionized. Specific conductance ≤10 μS/cm.
- 2. Sodium bicarbonate (NaHCO₃), reagent grade.
- 3. Chloroacetic acid, ≥99%.*
- 4. Eluent: 1.5 m <u>M</u> NaHCO₃. Dissolve 0.504 g NaHCO₃ in 4 L filtered, deionized water.
- Calibration stock solution, 1000 µg/mL. Dissolve 100 mg chloroacetic acid in 100 mL filtered, deionized water.
 - * See SPECIAL PRECAUTIONS.

EQUIPMENT:

- 1. Sampler: glass tube, 7 cm long, 6-mm OD, 4-mm ID, with plastic caps, containing two sections of 20/40 mesh silica gel (front = 100 mg; back = 50 mg) contained and separated by three silanized glass wool plugs. Pressure drop across the tube at 0.2 L/min is ca. 0.6 kPa (2.6 in. H₂O). Tubes are commercially available (SKC, Inc. 226-47-01, or equivalent). NOTE: Chloroacetic acid is irreversibly adsorbed on urethane plugs. Use sorbent tubes with glass wool plugs.
- 2. Personal sampling pump, 0.05 to 0.2 L/min, with flexible connecting tubing.
- 3. Ion chromatograph (IC), anion separator and guard column, anion suppressor (page 2008-1), conductivity detector, integrator, and strip chart recorder.
- 4. Ultrasonic bath.
- 5. Vials, 20-mL, glass, with aluminum-lined plastic screw caps.
- 6. Syringes, 3-mL, polyethylene with luer tip.
- Filter holder, luer tip, 13-mm, with PTFE filter, 5-µm pore size, or PTFE syringe filter.
- 8. Pipets, 10-µL to 2-mL.
- 9. Flasks, volumetric, 10- and 100-mL.

SPECIAL PRECAUTIONS: Chloroacetic acid is irritating to skin and mucous membranes [4]. Work with the concentrated material only in a hood.

SAMPLING:

- 1. Calibrate each personal sampling pump with a representative sampler in line.
- 2. Break ends of sampler immediately before sampling. Attach sampler to personal sampling pump with flexible tubing.
- 3. Sample at an accurately known flow rate between 0.05 to 0.2 L/min for a total sample size of 1 to 100 L.
- 4. Cap the samplers. Pack securely for shipment. NOTE: Store samples in the dark. Refrigerate samples if stored longer than 7 days.

SAMPLE PREPARATION:

- 5. Allow refrigerated samples to equilibrate to room temperature.
- 6. Transfer front sorbent section with front glass wool plug to vial. Place back sorbent section and other two glass wool plugs in separate vial.
- 7. Add 2.0 mL deionized water to each vial. Cap immediately.
- 8. Agitate vials in ultrasonic bath for 30 min at room temperature.
- 9. Draw sample extract through 13-mm PTFE filter with 3-mL syringe.

CALIBRATION AND QUALITY CONTROL:

- 10. Calibrate daily with at least six working standards.
 - a. Add known aliquots of calibration stock solution to deionized water in 10-mL volumetric flasks and dilute to the mark. Use serial dilutions as needed to obtain chloroacetic acid concentrations in the range 0.02 to 40 µg/mL.
 - b. Analyze together with samples and blanks (steps 13 through 15).
 - c. Prepare calibration graph [peak height (mm or μ S) vs. μ g chloroacetic acid per sample].
- 11. Determine desorption efficiency (DE) for each batch of silica gel used for sampling in the calibration range. Prepare at least three tubes at each of five levels.
 - a. Place silica gel from unused front section in vial.
 - b. Inject a known amount (2 to 20 μ L) of calibration stock solution, or a serial dilution thereof, onto front sorbent section with a microliter syringe.
 - c. Cap the vial. Allow to stand overnight.
 - d. Desorb (steps 7 through 9) and analyze together with working standards (steps 13 through 15).
 - e. Prepare graph of DE vs. µg chloroacetic acid recovered.
- 12. Analyze three quality control spikes and three analyst spikes to ensure that the calibration graph and DE graph are in control.

MEASUREMENT:

- 13. Set ion chromatograph according to manufacturer's recommendations and to conditions given on page 2008-1.
- 14. Inject sample aliquot manually or use autosampler.
 - a. Flush sample loop with 0.5 mL sample extract, then inject 0.5 mL sample.
 - b. Rinse sample loop with 1 to 2 mL deionized water between determinations of separate samples.

NOTE: All samples, eluents, and water flowing through the IC must be filtered to avoid plugging the system valves or columns.

15. Measure peak height.

NOTE: If sample peak height exceeds linear calibration range, dilute with deionized water, reanalyze, and apply appropriate dilution factor.

CALCULATIONS:

- 16. Determine mass, μg (corrected for DE), of analyte found in the sample front (W _f) and back (W_b) sorbent sections, and in the average media blank front (B _f) and back (B_b) sorbent sections.
- 17. Calculate concentration, C, of chloroacetic acid in the air volume sampled, V (L):

$$C = \frac{W_{f} + W_{b} - B_{f} - B_{b}}{V}, mg/m^{3}.$$

EVALUATION OF METHOD:

This method was developed and evaluated by Southern Research Institute [1] using dynamicallygenerated atmospheres of chloroacetic acid over the concentration range of 0.35 to 29 mg/m⁻³ at 25 to 27 °C and at relative humidity \geq 80%. Average recovery based on 18 samples, six at each of three levels, was 98% representing a negligible bias. Precision at 0.35 mg/m⁻³ was inhomogeneous with those of higher levels; therefore, precisions were not pooled. Using this poorest precision (S _r = 0.064), the overall precision (\hat{S}_{rT}) was estimated to be \leq 0.081. The breakthrough volume of the 100-mg sorbent section was found to be >100 L at 0.2 L/min when sampling chloroacetic acid concentrations of 60 mg/m⁻³ at 42 °C and RH of 10 and 80% and 35 mg/m⁻³ at 27 °C and 10 and 90% RH. Samples stored at ambient temperature for 7 days had a mean recovery of 91% and a precision, S_r, of 0.047. Samples refrigerated after day 7, and stored for 32 days exhibited a mean recovery of 100% with a precision, S_r, of 0.085 based on samples analyzed on day 1.

REFERENCES:

- [1] Dillon, H. K., D. W. Mason, and K. W. Boyd. Development of Air Sampling and Analytical Methods for Toxic Chlorinated Organic Compounds: Research Report for Monochloroacetic Acid, NIOSH Contract 210-78-0012, Southern Research Institute, Birmingham, AL (1980).
- [2] NIOSH Manual of Analytical Methods, 2nd ed., Vol. 6, P&CAM 322, U.S. Department of Health and Human Services, Publ. (NIOSH) 80-125 (1980).
- [3] McCullough, P. R. and J. W. Worley. "Sampling of Chloroacetyl Chloride in Air on Solid Support and Determination by Ion Chromatography," <u>Anal. Chem.</u>, <u>51</u>, 1120-1122 (1979).
- [4] Merck Index, 11th ed., Merck & Co., Rahway, NJ (1989).

METHOD REVISED BY:

Mary Ellen Cassinelli, NIOSH/DPSE.